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Unit -2
Plant Genetic Engineering
Confirmation of Transgenic plants

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Confirmation of Transgenic Plants

Confirmation of Transgenic Plants: A Detailed Note

The process of confirming the creation of transgenic plants involves several essential steps to ensure that the foreign genetic material (transgene) has been successfully introduced into the plant genome and is functioning as intended. Below is a detailed explanation of the various methods and techniques used to confirm transgenic plants.

1. Molecular Techniques for Confirmation

a. Polymerase Chain Reaction (PCR)

PCR is the most commonly used method for confirming the presence of a transgene in transgenic plants. This technique amplifies specific DNA sequences, allowing detection of the introduced gene.

- **Steps involved:**
 - Extract genomic DNA from the plant tissue.
 - Design primers that are complementary to the transgene sequence or its surrounding regions.
 - Perform PCR to amplify the target DNA sequence.
 - Analyze the PCR products using gel electrophoresis.
 - A positive result shows the expected band corresponding to the transgene.
- **Advantages:**
 - High sensitivity and specificity.
 - Can detect even small amounts of the transgene in plant tissues.

Confirmation of Transgenic Plants

b. Southern Blotting

Southern blotting is a technique used to detect the integration of the transgene into the plant genome. It provides information about the number of copies of the transgene and its integration pattern.

- **Steps involved:**

- Extract genomic DNA from the plant.
- Digest the DNA with restriction enzymes.
- Separate the DNA fragments by gel electrophoresis.
- Transfer the separated DNA to a membrane.
- Hybridize the membrane with a labeled probe complementary to the transgene.
- Visualize the bands using autoradiography or other detection methods.

- **Advantages:**

- Provides information on the size and integration pattern of the transgene.
- Can detect both the presence and number of transgene copies.

Confirmation of Transgenic Plants

c. Fluorescence in Situ Hybridization (FISH)

FISH is used to localize the transgene within the plant's chromosomes.

- **Steps involved:**

- Extract and prepare chromosome spreads from plant tissue.
- Hybridize the plant chromosomes with a fluorescently labeled probe specific to the transgene.
- Visualize the signal under a fluorescence microscope.

- **Advantages:**

- Allows precise localization of the transgene on chromosomes.
- Useful for determining if the transgene has integrated into specific chromosomal regions.

Confirmation of Transgenic Plants

2. Gene Expression Confirmation

a. Reverse Transcription PCR (RT-PCR)

RT-PCR is used to confirm that the transgene is not only present but is also being expressed in the plant.

- **Steps involved:**

- Extract RNA from the plant tissue.
- Convert RNA into complementary DNA (cDNA) using reverse transcriptase.
- Use PCR to amplify the cDNA corresponding to the transgene.
- Analyze the PCR products using gel electrophoresis.

- **Advantages:**

- Confirms the expression of the transgene at the mRNA level.
- Can be quantitative if performed as qRT-PCR.

Confirmation of Transgenic Plants

b. Northern Blotting

Northern blotting is used to detect and quantify specific RNA molecules, providing insight into the transcription of the transgene.

• **Steps involved:**

- Extract total RNA from plant tissues.
- Separate RNA by gel electrophoresis.
- Transfer to a membrane and hybridize with a labeled probe specific to the transgene's mRNA.
- Visualize the signals.

• **Advantages:**

- Allows detection of the transgene's transcript and provides quantitative information on its expression levels.

Confirmation of Transgenic Plants

C. Western Blotting

Western blotting detects and quantifies proteins expressed from the transgene, confirming its expression at the protein level.

- **Steps involved:**

- Extract protein from plant tissues.
- Separate proteins by gel electrophoresis.
- Transfer the proteins to a membrane.
- Use an antibody specific to the protein encoded by the transgene to detect its presence.
- Visualize the signal using chemiluminescence or other methods.

- **Advantages:**

- Confirms that the transgene product (protein) is being synthesized in the plant.
- Provides information about the size and abundance of the protein.

Confirmation of Transgenic Plants

3. Phenotypic Analysis

While molecular techniques provide direct evidence of the transgene, phenotypic analysis is also crucial in confirming the function of the transgene in the plant.

- **Steps involved:**

- Evaluate the physical and morphological traits of transgenic plants compared to non-transgenic (wild-type) controls.
- Assess any changes related to the transgene, such as resistance to pests, improved nutritional content, or altered growth patterns.
- For example, a transgenic plant with an insect-resistant gene may show increased resistance to specific pests.

- **Advantages:**

- Demonstrates the functional effect of the transgene.
- Useful for confirming traits like herbicide resistance, drought tolerance, or enhanced nutrient content.

Confirmation of Transgenic Plants

4. Protein-based Confirmation

In some cases, especially when dealing with transgenic plants designed to produce a foreign protein, confirmation of protein production is necessary.

- **Enzyme-linked Immunosorbent Assay (ELISA):** This method detects the presence of a specific protein (e.g., a foreign protein produced by the transgene) using antibodies.
- **Steps involved:**
 - Extract proteins from plant tissue.
 - Apply the extract to an ELISA plate that is coated with antibodies specific to the transgenic protein.
 - Add substrate and measure the color change, which indicates the amount of transgenic protein present.
- **Advantages:**
 - Highly specific for detecting the transgenic protein.
 - Quantitative and sensitive.

Confirmation of Transgenic Plants

5. Field Testing and Environmental Considerations

Once molecular confirmation and functional analysis are performed in the laboratory, field testing is often necessary to assess the stability and expression of the transgene in real-world conditions. This includes evaluating the plant's behavior across multiple generations, its resistance to environmental stress, and its potential impact on surrounding ecosystems.



Thank you